Chapter 01

Emergence of a New Paradigm in Understanding the Cardiovascular System: Pulse Synchronized Contractions

Mangel A* and Lothman K

RTI Health Solutions, USA

*Corresponding Author: Allen Mangel, RTI Health Solutions, 200 Park Offices Drive, PO Box 12194, Research Triangle Park, NC 27709-2194, USA, Tel: +1-919- 485-5668; Fax: +1-919-541-1275; Email: amangel@rti.org

First Published March 21, 2018

This Book Chapter is an excerpt from an article published by Mangel A and Lothman K at Cardiovascular Pharmacology: Open Access in September 2017. (Mangel A, Lothman K (2017) Emergence of a New Paradigm in Understanding the Cardiovascular System: Pulse Synchronized Contractions. Cardiovasc Pharm Open Access 6: 220. Doi: 10.4172/2329-6607.1000220)

Copyright: © 2018 Mangel A and Lothman K.

This article is distributed under the terms of the Creative Commons Attribution 4.0 International License

(http://creativecommons.org/licenses/by/4.o/), which permits unrestricted use, distribution, and reproduction in any medium, provided you give appropriate credit to the original author(s) and the source.

Keywords

Cardiovascular; Pulse Synchronized Contractions; Windkessel

What We Will Show

- The large conduit arteries undergo rhythmic smooth muscle activation in synchrony with the cardiac cycle.
- The contractions are neurogenic and are denoted as pulse synchronized contractions (PSCs).
- PSCs are not a movement artifact from the pulse wave or heartbeat.
- The pacemaker for the PSCs is in the right atrium.
- The smooth muscle wall of large arteries can contract as fast as the heartbeat.

What Was Believed in Gastrointestinal Smooth Muscle

An increase in intracellular calcium activates contractions in muscle cells. Because smooth muscle cells are long, narrow-diameter cells, it was believed that an influx of calcium could serve as the sole source of activator calcium for contractions following changes in membrane potential. Therefore, it was believed that no depolarization-mediated release of intracellularly stored calcium occurred. In a series of studies [1-3], we showed this not to be the case (Figures 1-3).

Rhythmic Membrane Potential Changes Normal saline 0 ---70 -- Calcium-free saline

Most Gastrointestinal Smooth Muscles Show

Figure 1: Slow waves with spikes (upper trace) are the recognized trigger for contractions in the gastrointestinal tract. Following incubation in calcium-free saline, an alternative rhythmicity develops (lower trace) [1,2].

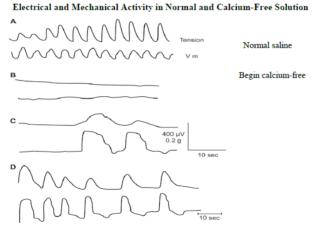


Figure 2: During incubation in calcium-free solution (beginning with Trace B), an alternative electrical activity with contractions develops [1,2]. Since contractions are observed in Traces C and D calcium release is occurring.

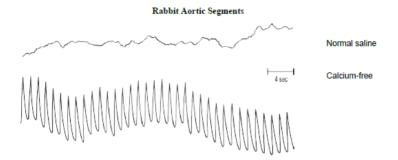


Figure 3: In contrast to gastrointestinal muscle segments, incubation of aortic segments from rabbits in normal saline is electrically quiescent (upper trace). In calciumfree solution, a fast rhythmic electrical event is produced (lower trace), but the muscle segments remain mechanically quiescent [3].

Windkessel Hypothesis: Otto Frank

- The prevailing hypothesis describing the behavior of the smooth muscle wall of the large arteries is that the wall does not contract in synchrony with the cardiac cycle but, rather, behaves as a passive elastic tube being rhythmically distended by pulsatile pressure changes. Neural input may modulate tone.
- Thus, it was believed that there was no vascular smooth muscle rhythmicity in synchrony with the cardiac cycle [4].

Proponents of the Windkessel Hypothesis Have Ignored

- Heyman, in a series of studies in man and dog, published between 1955 and 1961 [5-8], showed:
 - Extra-arterially recorded brachial pulses sometimes preceded intra-arterial pulses, suggesting arterial diameter varies in advance of pressure changes during the cardiac cycle.

- The difference between the extra-arterially recorded and intraarterially recorded pulse waves was abolished by stellate ganglion block, suggesting a neurally mediated event.
- It was concluded that: "the behaviour of the artery in the pulse is contradictory to principles of passive elasticity but seem to provide evidence of active participation of the arterial wall..."
- This series of papers has been largely ignored.

Hypothesis

Based on the ability of the aortic smooth muscle wall to generate fast rhythmic electrical activity in calcium-free solution (Figure 3), we sought to determine if the aortic smooth muscle wall could potentially show fast rhythmic contractile activity in vivo (Figures 4 and 5).

Methodology for in vivo Mechanical Activity Recording

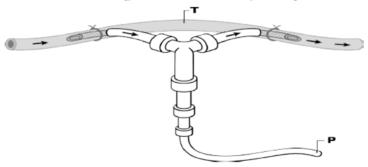


Figure 4: Recording technique for measurement of contractile activity in the in vivo rabbit aorta. Configuration represents a segment of aorta having blood flow bypassed and tension (T) recorded from the bypassed segment. Pulse pressure changes (P) were recorded from the non-bypassed segment [9,10].

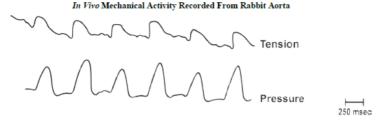


Figure 5: Using the recording technique shown in Figure 4, rhythmic tension changes (pulse synchronized contractions [PSCs]) were recorded with a 1:1 correspondence to the pulse wave [9-12].

Considerable Effort Was Expended Proving PSCs Were Not Due to a Mechanical Artifact

- Eliminate pulse wave (Figures 6 and 7)
- Eliminate cardiac contractility (Figures 6 and 7)
- Dispel prejudice that smooth muscle cannot "contract that fast" (Figure 8)

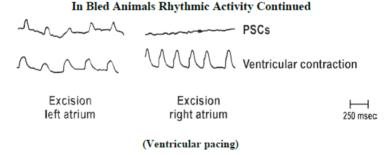


Figure 6: Following bleeding of rabbits, PSCs continued. In this configuration, ventricular muscle contractions were also recorded and pacing of the ventricles occurred. These studies (a) eliminated the pulse wave as an artifact, as animals were bled; (b) eliminated cardiac contractions as an artifact, as following excision of the right atrium with ventricular contractions paced to supra baseline levels, PSCs were not produced; and (c) suggested the PSC pacemaker is in the right atrium as excision of the right, but not left atrium, abolished PSCs [9].

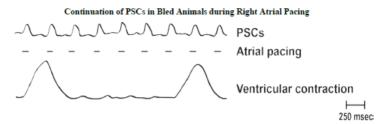


Figure 7: Shown above is an example of right atrial pacing in a bled rabbit. PSCs followed the pacing rate. In this and other animals, heart block developed with corresponding large amplitude ventricular contractions. This experiment supports both the pacemaker for PSCs residing in the right atrium and that PSCs are not secondary to a movement artifact from the heart [9].

Local Application of TTX on Electrically Stimulated Aortic Contractions

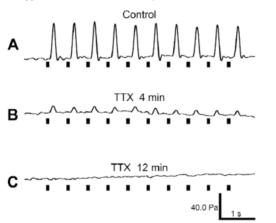


Figure 8: Electrical stimulation of the rat aorta *in vivo* produced contractions similar to PSCs. As PSCs are, these contractions were eliminated by the neural blocker tetrodotoxin (TTX). Black bars represent timing of stimulation [13].

Vessels Where PSCs Have Been Observed

| Species | Vessel |
|---------|-------------------------------------|
| Dog | Coronary, femoral, carotid arteries |
| Rabbit | Aorta |
| Cat | Pulmonary artery |
| Rat | Aorta |
| Human | Brachial artery |

From references [5-13]

PSCs

To evaluate whether the arterial smooth muscle wall is capable of contracting at the frequency of the heartbeat, electrical stimulation of the aorta *in vivo* was performed (Figure 8).

Conclusion

- The smooth muscle wall of the large arteries is capable of undergoing rapid contractions (PSCs) at the rate of the heartbeat.
- The contractions are neurogenic in origin as evidenced by blockade by TTX or lidocaine [references 9-13] and are not secondary to movement artifacts from the pulse wave or heartbeat.
- The pacemaker for the PSCs is in the right atrium.
- Direct electrical stimulation of the nerves within the aorta yields similar contractile activity.
- PSCs represent a modified platform to understand the etiology of cardiovascular diseases allowing for the development of new therapeutic targets.
- PSCs have been recently reviewed [14,15].

References

- Mangel AW, Nelson DO, Connor JA, Prosser CL. Contractions of cat small intestinal smooth muscle in calcium free solution. Nature. 1979; 281: 582-583.
- Mangel A, Nelson DO, Rabovsky JL, Prosser CL, Connor JA. Depolarization induced contractile activity of smooth muscle in calcium free solution. Am J Physiol. 1982; 242: 36-40.
- 3. Mangel A, van Breemen C. Rhythmic electrical activityin rabbit aorta induced by EGTA. J Exp Biol. 1981; 90: 339-342.
- 4. Frank O. The basic shape of the arterial pulse. First treatise: mathematical analysis. 1899. J Mol Cell Cardiol. 1990; 22: 255-277.
- 5. Heyman F. Movements of the arterial wall connected with auricle systole seen in cases of atrioventricular heart block. Acta Med Scand. 1955: 152: 91-96.
- 6. Heyman F. Comparison of intra-arterially and extra-arterially recorded pulse waves in man and dog. Acta Med Scan. 1957; 157: 503-510.
- 7. Heyman F. Extra- and intra-arterial records of pulse waves and locally introduced pressure waves. Acta Med Scan. 1959; 163: 473-475.
- 8. Heyman F. The arterial pulse as recorded longitudinally, radially and intra-arterially on the femoral artery of dogs. Acta Med Scan. 1961; 170: 77-81.
- 9. Mangel A, Fahim M, van Breemen C. Control of vascular contractility by the cardiac pacemaker. Science. 1982; 215: 1627-1629.
- 10. Mangel A, Fahim M, van Breemen C. Rhythmic contractile activity of the in vivo rabbit aorta. Nature. 1981; 289: 692-694.

- Mangel A, van Breemen C, Fahim M, Loutzenhiser R. Measurement of in vivo mechanical activity and extracellular CA45 exchange in arterial smooth muscle. In: Bevan JA, editor. Vascular Neuroeffector Mechanisms. 1983; 4: 347-351.
- 12. Ravi K, Fahim M. Rhythmic contractile activity of the pulmonary artery studied in vivo in cats. J Autonom Nerv Sys. 1987; 18: 33-37.
- Sahibzada N, Mangel AW, Tatge JE, Dretchen KL, Franz MR, et al. Rhythmic aortic contractions induced by electrical stimulation in vivo in the rat. PLoS One. 2015; 10: e0130255.
- 14. Mangel AW. Does the aortic smooth muscle wallundergo rhythmic contractions during the cardiac cycle. Exper Clinical Cardiol. 2014; 20: 6844-6851.
- 15. Mangel AW. A changing paradigm for understanding the behavior of the cardiovascular system. J Clin Exp Cardiol. 2017; 8: 496.